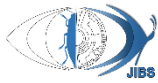


Original Article 

A new species of the genus *Aryenis* Bates, 1868 (Coleoptera: Tenebrionidae, Pimeliinae) from the Peruvian coastal desert

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ABSTRACT. A new species of tenebrionid beetle is described based on specimens collected in Peruvian coastal desert localities of Ancash, La Libertad, and Piura: *Aryenis yaserin* sp. nov. Type specimens and comparative material deposited in Peruvian entomological collections were the main basis for the study. Diagnosis, description, notes about biology, photos, and a distribution map are provided for this new species. Also, it is compared with the four previously described species in the genus *Aryenis* Bates, 1868, and a distribution map for the five known species is presented. The findings of the present work are discussed in relation to the current status of knowledge on systematics and biogeography of the genus *Aryenis*.

KEYWORDS: Darkling beetles, Evaniosomini, Arid zones, Neotropics, Systematics

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INTRODUCTION

The tribe Evaniosomini comprises seven Neotropical genera and one Afrotropical genus (Bouchard et al. 2021). Initially, the tribe only included *Evaniosomus* Guérin-Méneville, 1834; *Evelina* Thomson, 1860, and *Melaphorus* Guérin-Méneville, 1834 (Thomson 1860), to which *Aryenis* Bates, 1868, and *Chorasmius* Bates, 1868 were added (Bates 1868, 1872). Later, Gebien (1937) placed *Achanius* Erichson, 1847 in Evaniosomini, and it was treated as such by Kulzer (1950, 1956), who also described the new genus *Vaniosus* Kulzer, 1956. Finally, *Oppenheimeria* Koch, 1952 was transferred from Epitragini to Evaniosomini (Endrödy-Younga 1998). Cladistic and phylogenomic analyses performed for Pimeliinae genera support the close relationship between *Aryenis*, *Evaniosomus*, and *Melaphorus* as the core of the Evaniosomini tribe, and on the other hand, the placement of *Achanius* in the Edrotini tribe (= Eurymetopini) (Doyen, 1993; Ragionieri et al. 2023). Nevertheless, Flores & Aballay (2015) suggested the inclusion of *Achanius* and *Vaniosus* in Evaniosomini, based on character states of mandibles and the internal female reproductive tract. The relationships of *Chorasmius*, *Evelina*, and *Oppenheimeria* with other genera proposed for Evaniosomini remain largely unresolved.

Species in the genus *Aryenis* have been described from scattered localities in South America over a 150-year interval (Blanchard 1843; Bates 1868, 1872; Fairmaire 1892; Peña 1994): *A. unicolor* (Blanchard, 1843) from Bolivia, *A. rufescens* Bates, 1868 from Argentina, *A. haagi* Bates, 1872 from Peru, *A. minor* Fairmaire, 1892 from Argentina and *A. tenuis* Peña, 1994 from Chile. Following Bates's (1868) suggestion, the first two species were subsequently treated as synonyms (Gebien 1937; Blackwelder 1945), and the redefined *A. unicolor* was later widely recorded in Argentina and also in Chile (Peña 1961). Until now, only *A. haagi* has been recorded in Peru, specifically in coastal deserts (0–1000 m) located west of the Andes (Smith et al. 2015; Giraldo & Flores 2016).

The purposes of the present work are to describe a new Peruvian species of the genus *Aryenis* and compare it with the four previously described species of the genus.

MATERIAL AND METHODS

This study was carried out with the collaboration of the following institutions: Natural History Museum, London, United Kingdom (NHMUK); Instituto Argentino de Investigaciones de las Zonas Áridas, Mendoza, Argentina (IADIZA); Museo de Entomología Klaus Raven Büller, Universidad Nacional Agraria La Molina, Lima, Peru (MEKRB); Museo de Historia Natural Javier Prado, Universidad Nacional Mayor de San Marcos, Lima, Peru (MUSM). Relevant morphological characters for Evanosomini genera and *Aryenis* species were obtained from descriptions and images available in Bates (1868, 1872), Peña (1994), and Flores & Aballay (2015). The male genitalia of the new species are not figured because comparative images of genitalia are required for all species, but the author lacks material for some taxa. The species *A. haagi* and *A. rufescens* were studied from photos of holotypes (NHMUK) and comparative material housed in Peruvian collections (MEKRB, MUSM). Type specimens were indicated by red and yellow printed labels, respectively, bearing the status of the specimens, the name of the species, the name of the author, and the year of the designation.

Specimens of the new species were photographed with a Canon® EOS Rebel T5i DSLR, equipped with a macro lens. Photos were edited using Combine ZP (Hadley 2006) and graphic design software. Distribution maps were elaborated using SimpleMappr (Shorthouse 2010).

RESULTS

Order Coleoptera Linnaeus, 1758

Family Tenebrionidae Latreille, 1802

Subfamily Pimeliinae Latreille, 1802

Tribe Evanosomini Lacordaire, 1852

Genus *Aryenis* Bates, 1868

Type species. *Aryenis rufescens* Bates, 1868 (= *Statira unicolor* Blanchard, 1843)

Aryenis yaserin Giraldo sp. nov.

<https://zoobank.org/urn:lsid:zoobank.org:act:C802392F-5B72-40C5-9E0E-F005A7136938>

[Figs 1A–D, 2A–B]

Material examined. **Holotype** ♂ (MUSM), Peru, La Libertad, Virú, Chao, 08°27'07.97"S 78°32'21.13"W, 440 m, 13-X-2021, N. Zenteno leg [*Aryenis yaserin* sp. nov./HOLOTYPUS/Det. A. Giraldo 2025] (Fig. 1A); **Paratypes** (IADIZA, MEKRB, MUSM): 1♂, 1♀ (IADIZA), 2♀♀ (MUSM), same data as holotype; 1♂ (MUSM), Peru, La Libertad, Virú, Chao, 100 m, nocturno, 12-III-1993, P. Hocking leg; 1♀ (MUSM), Peru, Ancash, Santa, Chimbote, 09°07'0.1"S, 78°33'17.20"W, 0 m, 01-VIII-2023, M. Basurto leg; 1♀ (MUSM), Peru, Piura, Talara, Los Órganos, 04°13'02.4"S 81°01'42.9"W, 260 m, 15-III-2014, L. Figueroa leg; 2 not sexed (MEKRB), Peru Piura, Sullana, Marcavelica, Samán, 04°50'12.77"S 80°45'49.73"W, 50 m, 16-XI-1989, P. Castillo leg [*Aryenis yaserin* sp. nov./PARATYPUS/Det. A. Giraldo 2025].

Additional materials. **Holotype** of *Aryenis haagi* Bates, 1872 (NHMUK), [Type] [PERU] [*aryenis*/Haagi/F. Bates/type] [F. Bates./81-19.] [NHMUK015019600] (Fig. 1E); **Holotype** of *Aryenis rufescens* Bates, 1872 (NHMUK), [Type] [ARGNT/NE REP] [*aryenis/rufescens*/type. F. Bates] [F. Bates./81-19.] [NHMUK 015020339] (Fig. 1F), studied from photos of dorsal aspects provided by courtesy of Dmitry Telnov; **Other specimens** of *Aryenis haagi* Bates, 1872, 2♂♂ (MEKRB), Peru, Lima, Huaral, Iguanil, 11°24'29.87"S 77°12'51.87"W, 400 m, II-2000, W. Tori leg; 1 not sexed (MUSM), Peru, Lima, Huaral, near to Reserva Nacional de Lachay, 11°23'14"S 77°17'06"W, 417 m, 01-04-III-2019, M. Lozano leg; 1 not sexed (MUSM), Peru, Lima, Huarochirí, San Antonio, 11°51'27.79"S 76°38'17.24"W, 1366 m, 20-28-III-2019, E. Gamboa leg; 7♂♂ (MEKRB), Peru, Lima, Huaura, Reserva Nacional de Lachay, 11°21'53.93"S 77°22'33.80"W, 400 m, 04-VI-2018, A. Giraldo leg.

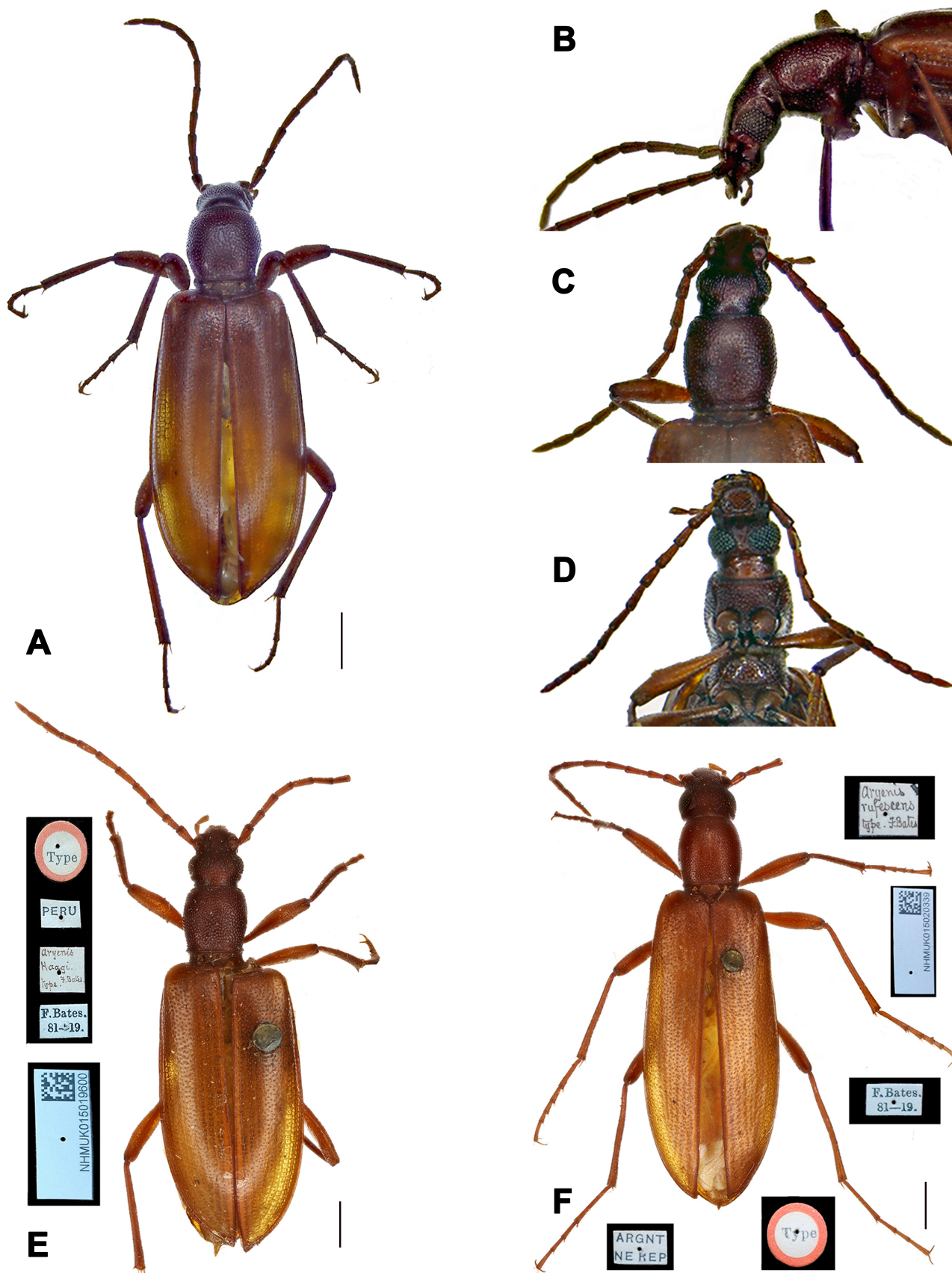


Figure 1. *Aryenis yaserin* Giraldo sp. nov., holotype (MUSM): **A.** Dorsal habitus, **B.** Head and pronotum, lateral view; female paratype (MUSM): **C.** Head and pronotum, dorsal view, **D.** Head, prosternum and mesosternum, ventral view; *Aryenis haagi* Bates, 1872, holotype (NHMUK): **E.** Dorsal habitus; *Aryenis rufescens* Bates, 1868, holotype (NHMUK): **F.** Dorsal habitus. Photos of NHMUK type specimens provided by Dmitry Telnov. Scale bars = 1 mm.

Etymology. The specific name is an anagram of the generic name, alternating consonants in reverse order and vowels in sequential order.

Diagnosis. Body shiny, mostly dark ferrugineous, yellowish on elytra, metasternum, metepimeron, and abdominal ventrites. Frontal punctures dense, separated by one puncture diameter, forming an elongate-rugose design. Eyes dorsally separated by a distance equal to 3.0 eye diameters, ventrally separated by a distance equal to 0.25 eye diameters. Antennae with antennomeres 8–10 parallel-sided. Pronotal punctures dense, separated by a distance equal to 1.0–1.5 puncture diameters. Elytral striae composed of dense puncture series, separated by a distance equal to 1.0 puncture diameter. Tibiae nearly straight.

Description. — Holotype ♂, paratypes ♂ ♀. Length. 7.0–8.5 mm. Body elongate (length/width ratio = 2.7), slender, and dorsally flattened; shiny, head, pronotum, scutellar shield, antennae, legs, prosternum and mesoventrite dark ferrugineous, elytra, metaventrite, metepimeron and abdominal ventrites yellowish (Fig. 1A). Sexual dimorphism is inconspicuous at level of external morphology.

Head (Fig. 1B, C, D). Labrum hidden under epistome; epistome elongate, with anterior margin semicircular, nearly at same level with antennal insertions; epicanthi axe-shaped, with approximate same size than eye diameter; two basal thirds of epistome and frons with coarse and dense punctures, separated by one puncture diameter, punctures elongate-rugose on frons, punctures becoming fine and sparse on apical third of epistome and vertex. Eyes large, coarsely faceted; slightly convex dorsally, anterior and posterior margins parallel-sided in lateral view, ventrally close to each other; dorsal interocular distance equal to 3.0 eye diameters, ventral interocular distance equal to 0.25 eye diameters. Antennae elongate, slender, antennomeres 7–11 surpassing posterior margin of pronotum; antennomeres gradually becoming more parallel-sided from base to apex; antennomere 1 robust, antennomere 3 longer than 2 or 4 separately, antennomeres 5–10 longer than wide, antennomere 11 slightly shorter than 10, apically attenuated and somewhat pointed; antennomeres with coarse punctures and covered with minute golden setae, setae denser on antennomeres 8–11. Mentum transverse, pentagonal; confluence between gula and genae marked with a circular fossa; mandibles robust, base with nearly the same width than apex, outer margin rounded, inner margin straight, with a sharp apex and a notorious cusp on apical third; maxillar palpi conspicuous, apical maxillar palpomere drop-shaped; labial palpi shortened, hidden under mentum, as well as cardo and stipes.

Prothorax (Fig. 1B, C). Subcylindrical (length/width ratio = 1.05), widest at middle, half as wide as elytra. Anterior and posterior margins with thickened beads, bead of posterior margin constricts the pronotum just anterad its base; lateral margins not beaded; anterior angles obtuse and blunt, sloping towards venter; posterior angles obtuse and blunt. Punctures dense, separated by a distance equal to 1.0–1.5 puncture diameters. Prothoracic hypomeron contiguous to pronotum, with the same punctuation pattern. Prosternum strongly convex ventrally; with anterior margin beaded and laterally compressed apical tooth, which is also sharp and recurved; with coalescent punctures.

Pterothorax (Fig. 1D). Scutellar shield small, triangular, smooth, with margins thickened on apical half. Elytra elongate, slender, flattened on disc, with developed humeri; in dorsal view parallel-sided, gradually contracted on apical fifth; in lateral view not suddenly declivous apically; without dorsal carinae, with transverse basal carinae from sutura to each humeri; sutura and elytral lateral margins carinate throughout their lengths. Striae marked with series of minute punctures and fuzzy black spots, separated by a distance equals to 1.0 puncture diameter; intervals marked with a series of minute punctures, same sized than punctures on striae but fainter and sparser. Mesoventrite short; metepimeron elongate; metaventrite elongate, twice as long as mesoventrite. mesoventrite and metepimeron with punctures separated by a distance equal to one puncture diameter, metaventrite with punctures separated by a distance equals to 1.5 puncture diameters. Metathoracic wings developed.

Abdomen. Ventrites sparsely and faintly punctured.

Legs. Pro-, meso- and metacoxae pairs nearly contiguous; pro- and mesocoxae circular, metacoxae elongate oval, and transverse; distance between meso and metacoxae equals to 1.5 metacoxal individual length. Femora apically dilated, clavate. Tibiae nearly straight; coarsely punctured and covered with minute scattered and decumbent golden setae. Tarsi ventrally covered with minute spiny setae; basal protarsomeres subequal in length to apical ones; basal meso- and metatarsomeres 1.5 times as long as apical ones.

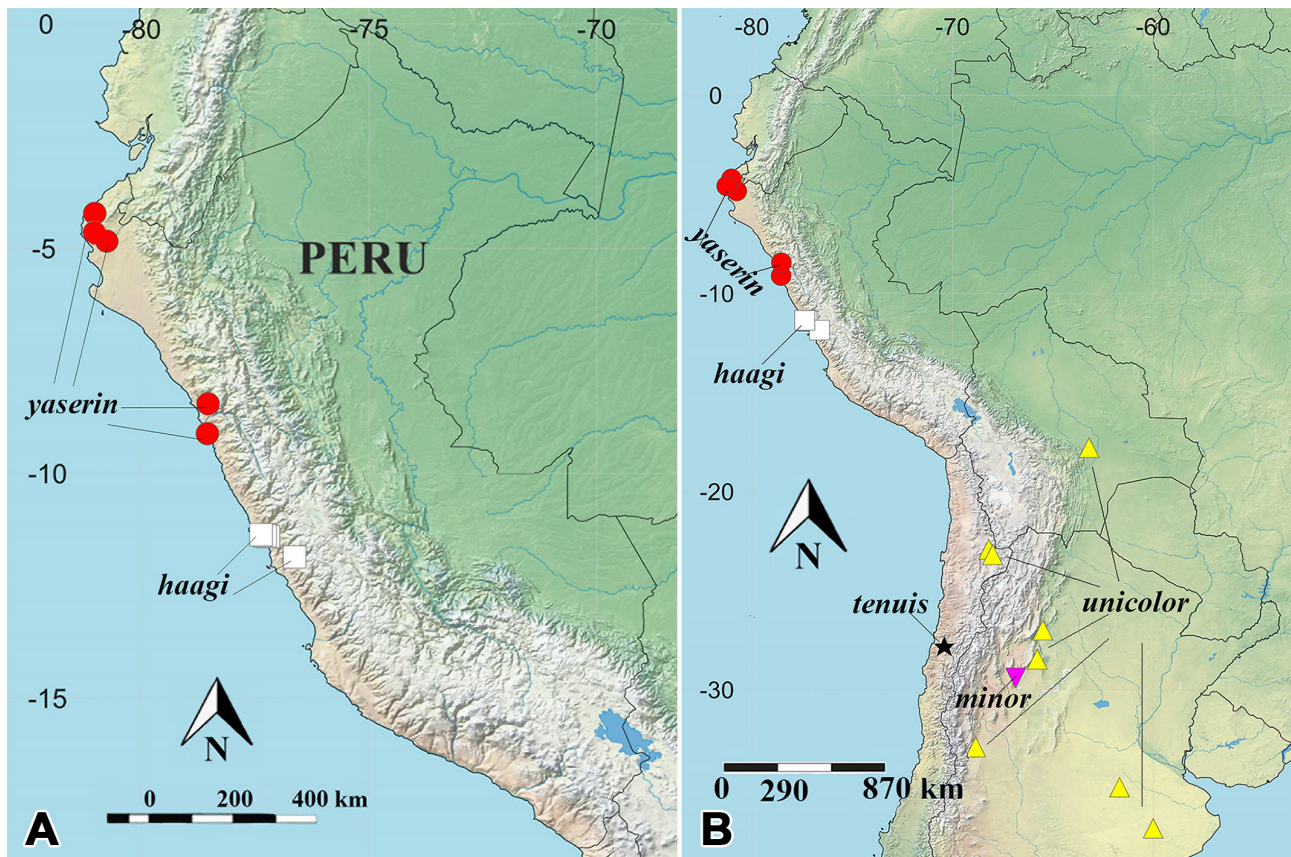


Figure 2. Distribution records of the *Aryenis* species, **A.** *Aryenis yaserin* Giraldo sp. nov., and *Aryenis haagi* Bates, 1872 in Peru, **B.** Five known species in South America.

Comparative Diagnosis. The new species is most similar to *A. haagi*, but it is slightly longer (8.5–10.0 mm); darker and duller (Fig. 1E); with ventrally nearly contiguous eyes separated by an interocular distance equal to 0.13 eye diameters and coalescent pronotal punctures separated by 0.0–0.5 puncture diameters. The differences are most obvious with *A. unicolor* (= *rufescens*), which is even longer (9–10 mm) (Fig. 1F; Blanchard 1843, Pl. XV, fig. 2); also has slender antennae (Bates 1868, fig. 1a); dorsally bulging eyes; ventrally distant eyes separated by an interocular distance equals to 0.5 eye diameters (Bates 1868, fig. 1b); sparse pronotal punctures separated by 1.0–2.0 puncture diameters and metatibiae obliquely bent on apical third. Using the original descriptions and illustrations, differences can be established between the two remaining species. *A. minor* and *A. tenuis* are smaller and paler species, with a peculiar shape of antennae; in *A. minor*, antennomere 2 is noticeably short, with lateral margins rounded (Peña 1994, fig. 2), while in *A. tenuis*, antennomeres 2, 3, and 11 are noticeably short, with lateral margins rounded (Peña 1994, fig. 1).

Biology and distribution. Little is known about its biology, except that it is a nocturnal flyer based on label data of a specimen collected at Chao. This trait may partly explain the scarcity of specimens in collections, since pitfall traps and direct searching have been preferred as sampling methods over light traps in the Peruvian coastal desert. The known distribution of this species ranges from Piura (4°S) to Ancash (9°S) in areas of coastal desert and seasonally dry forest (Fig. 2A). Undeterminate specimens of the genus *Aryenis* recorded by Juárez-Noe & González-Coronado (2019) from Talara (Piura), most probably belong to this new species.

DISCUSSION

The present contribution adds one more species to the genus *Aryenis*, tracing the road for its systematic revision, in which the known species could be reevaluated using a uniform set of morphological

characters. A study of this kind would be useful to support, for instance, the synonymy of *A. rufescens* under *A. unicolor*, cautiously suggested by Bates (1868) “It is just possible the above species may be identical with the *Statira unicolor* (Blanch. in d’Orbig. Voy. Amer. merid. p. 199), but his description is too short and vague to enable me to decide this point; judging from the figure given in his work...”, which was uncritically assumed in subsequent checklists (Gebien 1937; Blackwelder 1945). For this purpose, as many type specimens as possible should be examined, and new collecting trips should be conducted in suitable habitats, using light traps as the preferred sampling method.

As Flores & Pizarro-Araya (2006) noted, *Aryenis* is one of the thirteen South American genera of Pimeliinae widely distributed on both sides of the Andes, with one species recorded on both sides of this mountain range and the remaining species restricted either to the west or the east of it. Indeed, the distribution of the genus *Aryenis* covers several scattered locations in South America (Fig. 2B). The Peruvian species are known from the northern half of the country, with *A. yaserin* ranging from Piura to Ancash and *A. haagi* restricted to Lima (label data cited herein). Whereas *A. unicolor* (= *A. rufescens*) has the widest distribution, including Bolivia (Santa Cruz), Chile (San Pedro de Atacama, Toconao), and Argentina (Buenos Aires, Catamarca, Mendoza, Tucumán) (Peña 1961). In contrast, *A. minor* and *A. tenuis* are only known from their type localities, Argentina (La Rioja) (Fairmaire 1892) and Chile (Atacama) (Peña 1994), respectively.

AUTHOR’S CONTRIBUTION

The author confirms his contribution to the whole processing steps in the research, examination of the specimens, illustrations, and preparation of the manuscript. He read and approved the final version of the manuscript.

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This research received no specific grant from any funding agencies.

AVAILABILITY OF DATA AND MATERIAL

The specimens listed in this study are deposited in entomological collections of IADIZA, MEKRB, and MUSM and are available from their respective curators upon request.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

This study only included arthropod material, and all required ethical guidelines for the treatment and use of animals were strictly adhered to in accordance with international, national, and institutional regulations. No human participants were involved in any studies conducted by the authors for this article.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTERESTS

The authors declare that there is no conflict of interest regarding the publication of this paper.

GENERATIVE AI STATEMENT

The author declares not having used any kind of AI tools in the preparation of the manuscript.

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REFERENCES

- Bates, F. (1868) XVII. Descriptions of new genera and species of Heteromera. *Transactions of the Entomological Society of London*, 16, 309–326. <https://doi.org/10.1111/j.1365-2311.1868.tb00630.x>
- Bates, F. (1872) Notes on Heteromera, and descriptions of new genera and species (No. 2). *The Entomologist's Monthly Magazine*, 9, 133–135. <https://doi.org/10.5962/bhl.part.4726>
- Blackwelder, R.E. (1945) Checklist of the Coleopterous insects of Mexico, Central America, the West Indies and South America. Part 3. *Bulletin of the United States National Museum*, 185, 343–550. <https://doi.org/10.5479/si.03629236.185.3>
- Blanchard, C.É. (1843) Insectes de l'Amérique Méridionale recueillis par M. Alcide d'Orbigny. In: Bertrand, P. & Levrault V. (eds.) *Voyage dans l'Amérique Méridionale. Tome sixième. 2.e Partie: Insectes*. Paris & Strasbourg, pp. 214–216.
- Bouchard, P., Bousquet, Y., Aalbu, R.L., Alonso-Zarazaga, M.A., Merkl, O. & Davies, A.E. (2021) Review of genus-group names in the family Tenebrionidae (Insecta, Coleoptera). *Zookeys*, 1050, 1–633. <https://doi.org/10.3897/zookeys.1050.64217>
- Doyen, J.T. (1993) Cladistic relationships among Pimeliine Tenebrionidae (Coleoptera). *Journal of the New York Entomological Society*, 101(4), 443–514.
- Endrödy-Younga, S. (1998) The American tribe Evaniosomini in southern Africa (Coleoptera: Tenebrionidae). *Annals of the Transvaal Museum*, 36 (31), 425–426.
- Fairmaire, M.L. (1892) Descriptions de quelques coleopteres Argentins. *Annales de la Société Entomologique de Belgique*, 36, 242–253.
- Flores, G.E. & Aballay, F.H. (2015) Two Evaniosomini species (Coleoptera: Tenebrionidae) associated with decaying carcasses in Argentina, with remarks on the tribal assignment of *Achanius* Erichson. *The Coleopterists Bulletin*, 69 (14), 167–179. <https://doi.org/10.1649/0010-065X-69.mo4.167>
- Flores, G.E. & Pizarro-Araya, J. (2006) The Andes mountain range uplift as a vicariant event in the Pimeliinae (Coleoptera: Tenebrionidae) in southern South America. *Cahiers scientifiques*, 10, 95–102. <https://doi.org/10.3406/mhnl.2006.1352>
- Gebien, H. (1937) Katalog der Tenebrioniden. Teil I. *Pubblicazioni del Museo Entomologico Pietro Rossi. Duino*, 2, 505–883.
- Giraldo, A.E. & Flores, G.E. (2016) Peruvian Tenebrionidae: A review of present knowledge and biodiversity. *Annales Zoologici*, 66 (4), 499–513. <https://doi.org/10.3161/00034541ANZ2016.66.4.002>
- Hadley, A. (2006) CombineZM public domain image processing software. Available from: <https://combinezp.software.informer.com> [Accessed November 11, 2025]
- Juarez-Noe, G. & Gonzalez-Coronado, U. (2019) Actualización a la lista de coleópteros (Insecta: Coleoptera) de la Región Piura, Peru. *Graellsia*, 75 (2), e097. <https://doi.org/10.3989/graellsia.2019.v75.233>
- Kulzer, H. (1950) 3 Beitrag zur Kenntnis der Tenebrioniden. *Entomologische Arbeiten aus dem Museum Georg Frey*, 1, 9–46.
- Kulzer, H. (1956) Neue Tenebrioniden aus Südamerika. 15 Beitrag zur Kenntnis der Tenebrioniden. *Entomologische Arbeiten aus dem Museum Georg Frey*, 7, 895–962.
- Peña, L.E. (1961) Nuevos insectos para Chile (Coleoptera-Tenebrionidae). *Noticiario Mensual del Museo Nacional de Historia Natural*, 60, 1–6.
- Peña, L.E. (1994) Nuevas especies de Tenebrionidae (Insecta – Coleoptera) de la Región Neotropical. *Gayana Zoologia*, 58 (2), 151–168.
- Ragioneri, L., Zuñiga-Reinoso, A., Bläser, M. & Predel, R. (2023) Phylogenomics of darkling beetles (Coleoptera: Tenebrionidae) from the Atacama Desert. *PeerJ*, 11, e14848. <https://doi.org/10.7717/peerj.14848>
- Shorthouse, D. (2010) SimpleMappr, an online tool to produce publication-quality point maps. Available from: <http://www.simplemappr.net> [Accessed November 17, 2025]
- Smith, A.D., Giraldo, A.E., Flores, G.E. & Aalbu, R.L. (2015) Beetles (Coleoptera) of Peru: A survey of the families. Tenebrionidae. *Journal of the Kansas Entomological Society*, 88 (2), 221–228. <https://doi.org/10.2317/kent-88-02-221-228.1>
- Thomson, J. (1860) Evaniosomitarum enumeratio. *Musée Scientifique ou Recueil d'Histoire Naturelle*, 1, 21–23. [in Latin]

یک گونه جدید از جنس *Aryenis* Bates, 1868 (Coleoptera: Tenebrionidae, Pimeliinae) از بیابان ساحلی پرو

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چکیده: یک گونه جدید از سوسک‌های تاریک‌زی به نام *Aryenis yaserin* sp. nov. بر اساس نمونه‌هایی که از مناطق بیابانی ساحلی پرو در آنکاش، لالیبرتاد و پیورا جمع‌آوری شده بودند، توصیف شد. نمونه‌ها در مجموعه‌های حشره‌شناسی پرو نگهداری می‌شوند که پایه اصلی این مطالعه بوده‌اند. تشخیص، توصیف، یادداشت‌هایی درباره زیست‌شناسی، عکس‌ها و دامنه انتشار این گونه جدید ارائه شد. همچنین، این گونه با چهار گونه قبلاً توصیف شده در جنس *Aryenis* Bates, 1868 مقایسه شده و نقشه انتشار پنج گونه شناخته شده ارائه شد. یافته‌های این تحقیق در ارتباط با وضعیت کنونی دانش در زمینه سیستماتیک و زیست‌جغرافیای جنس *Aryenis* مورد بحث قرار گرفت.

ویراستار علمی

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واژگان کلیدی: سوسک‌های تاریک‌زی، Evaniosomini، مناطق خشک، نئوتروپیکال، سیستماتیک