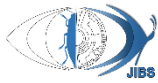


Short Article 

# Discovery of the genus *Clanoneurum* Becker, 1903 (Diptera: Ephydriidae) in Iran, representing an emerging pest of sugar beet, *Clanoneurum cimiciforme* (Haliday)

Ebrahim Gilasian<sup>1</sup>  | Elnaz Hamidi<sup>2</sup> 

1 Insect Taxonomy Research Department, Iranian Research Institute of Plant Protection (IRIPP), Agricultural Research, Education and Extension Organization (AREEO), 19395-1454, Tehran, Iran

2 Plant Protection Research Department, North Khorasan Agricultural and Natural Resources Research and Education Center, Agricultural Research, Education and Extension Organization (AREEO), Bojnord, Iran. Email: e.hamidi@areeo.ac.ir

Corresponding author: Ebrahim Gilasian | ✉ [ebrahimgilasian@gmail.com](mailto:ebrahimgilasian@gmail.com); [gilasian@iripp.ir](mailto:gilasian@iripp.ir)

<https://zoobank.org/urn:lsid:zoobank.org:6ADA951B-A8BF-4561-87B0-EFCF75D71149>

**Academic Editor**

Tadeusz Zatwarnicki

**Received**

February 23, 2026

**Revised**

April 22, 2026

**Accepted**

April 29, 2026

**Published online**

May 10, 2026

**ABSTRACT.** The genus *Clanoneurum* Becker, 1903 currently includes two valid species and is distributed across the Afrotropical, Nearctic, Oriental, and Palaearctic regions. This genus and its species, *C. cimiciforme* (Haliday, 1855) are reported for the first time from Iran (North Khorasan Province) as a leaf miner of sugar beet (*Beta vulgaris* L.). This species has also been collected from the mangrove habitats in southern Iran (Bushehr and Hormozgan Provinces) close to the Persian Gulf and the Oman Sea. The morphological characters of the genus and species, along with photographs of adults, immature stages, and symptoms of damage on sugar beet leaves, are presented.

**KEYWORDS:** *Beta vulgaris*, leaf miner, mangrove habitat, new record, Psilopini, Shore flies

**Citation:** Gilasian, E. & Hamidi, E. (2026) Discovery of the genus *Clanoneurum* Becker, 1903 (Diptera: Ephydriidae) in Iran, representing an emerging pest of sugar beet, *Clanoneurum cimiciforme* (Haliday). *Journal of Insect Biodiversity and Systematics*, 12 (03), 469–476.

## INTRODUCTION

Sugar beet (*Beta vulgaris* L.) is one of the principal sources of sucrose worldwide and has been cultivated commercially since the eighteenth century (Izzatullayeva et al. 2014). Its cultivation expanded markedly during the twentieth century and is currently practiced in approximately 50 countries across the world. In arable farming systems, sugar beet is commonly grown within three-year crop rotations, typically in combination with cereals such as winter wheat and winter barley. It may also be included in rotations with maize, common bean, potato, or oilseed rape (Märländer et al. 2003; Koga 2008). Although production is primarily concentrated in temperate regions of the Northern Hemisphere, recent advances in agricultural technologies and plant breeding have enabled its winter cultivation in certain tropical areas as well (Sharma et al. 2017).

Several families within the order Diptera include species whose larvae act as leaf miners in sugar beet cropping systems. Among these, the family Anthomyiidae has received the greatest research attention. Leaf-mining species have also been reported from the families Ephydriidae, Drosophilidae, and Agromyzidae (Hespenheide 1991). Within Anthomyiidae, members of the genus *Pegomya* Robineau-Desvoidy, 1830 are recognized as the most important leaf miners of sugar beet. However, in Iran, only *Pegomya hyoscyami* (Panzer, 1809) has been reported to date (Pour Rahim et al. 2016).

The family Ephydriidae (shore flies), comprising approximately 2,000 species worldwide (Mathis & Zatwarnicki 1995), plays important ecological roles, functioning both as minor pests and as a food resource for wildlife. Despite their rich diversity, shore flies in the Middle East remain insufficiently studied. Adults can be readily identified based on several diagnostic characteristics: a distinctly convex and protruding face; an unusually large oral opening; the wing venation: two costal breaks and the absence of the anal cell (Mathis & Zatwarnicki 1998). Larvae are typically aquatic or semi-aquatic and often exhibit morphological or physiological adaptations corresponding to their specific habitats. Nevertheless, certain genera inhabiting arid desert regions have been recorded at considerable distances from any visible water source (Dawah et al. 2019).

The genus *Clanoneurum* Becker, 1903 (Ephydriidae: Discomyzinae) comprises two valid species and exhibits a distribution spanning the Afrotropical, Nearctic, Oriental, and Palearctic regions. The species *C. cimiciforme* (Haliday, 1855) is primarily a Palearctic species, but its range extends to the Afrotropical and Oriental regions. As with the second member of this genus, this species appears to be associated with plants of the family Chenopodiaceae. Although the morphology of the immature stages and the biology of this species remain mostly unknown, its immature stage illustrations and the preliminary biological observations of *C. americanum* have been documented and examined by de Meijere (1947) and Wheeler (1982). However, there have been some studies conducted in recent years reporting this species from regions close to Iran. Mathis et al. (2017) provided identification keys to the genera of Ephydriidae, the tribe Psilopini, and also presented the diagnostic characteristics of *C. cimiciforme* as a new record from the United Arab Emirates. In their study of the family Ephydriidae, Dawah et al. (2019) reported this species from Saudi Arabia.

The aim of this study is to report the genus *Clanoneurum* and the species *C. cimiciforme*, for the first time from Iran, emphasizing that the larvae of this species should be recognized as an emerging leaf-miner pest in Iran and added to the list of known sugar beet pests in the country.

## MATERIAL AND METHODS

Leaves of sugar beet infested by *C. cimiciforme* larvae were sampled in the summer of 2025 from Hesar Garmkhan, Bojnord, North Khorasan Province (northeastern Iran) (Fig. 1). The samples were transferred to insect rearing containers and maintained at room temperature until the emergence of adults. The adult flies were preserved in 75% ethanol and sent to the Insect Taxonomy Research Department in Tehran for identification. Other specimens were collected from the mangrove forests in southern Iran using yellow pan traps (Fig. 4).

To prepare the specimens and prevent shrinkage after ethanol preservation, we followed the AXA method proposed by van Achterberg (2009). The specimens were subsequently mounted on cardboard cards, and labels indicating the collection site were affixed. The field photographs were taken using a mobile phone, Samsung A50, while adult specimens were photographed in the laboratory using a Canon 650D camera. All specimens are deposited in the Hayk Mirzayans Insect Museum [HMIM], Insect Taxonomy Research Department, Tehran. Accurate genus and species identification was conducted using reliable taxonomic literature. The morphological terminology used in this paper follows Mathis et al. (2021).

## RESULTS

**Class Insecta Linnaeus, 1758**

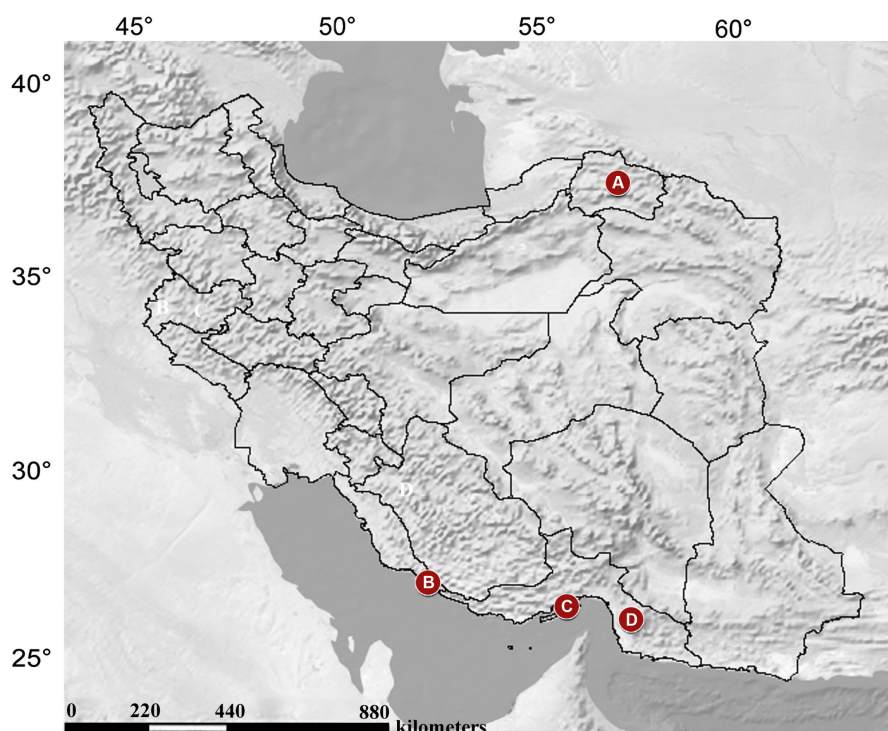
**Order Diptera Linnaeus, 1758**

**Suborder Brachycera Zetterstedt, 1842**

**Family Ephydriidae Zetterstedt, 1837**

**Subfamily Hydrelliinae Robineau-Desvoidy, 1830**

**Tribe Psilopini Cresson, 1925**



**Figure 1.** Distribution of *Clanoneurum cimiciforme* in Iran. **A.** Bojnord, Hesar-e Garmkhan (North Khorasan Province); **B.** Nayband (Bushehr Province); **C.** Qeshm Island (Hormozgan Province); **D.** Sirik (Hormozgan Province).

### Genus *Clanoneurum* Becker, 1903

*Clanoneurum* Becker, 1903:165. **Type species.** *Clanoneurum infumatum* Becker, 1903 [= *Discomyza cimiciforme* Haliday, 1855], by monotypy.

**Diagnosis.** Face flat and wrinkled; antenna shorter than eye; antennal base arising from lower part of head; scape oriented anteriorly, with a slight ventral inclination; pedicel and postpedicel similarly oriented ventrally, forming an obtuse angle with scape; postpedicel approximately twice as long as its height; reclinate and proclinate fronto-orbital setae present; wing venation distinctive:  $R_{2+3}$  runs along the Costa; discal medial crossvein (dm-m) with sharp angle medially.

### *Clanoneurum cimiciforme* (Haliday, 1855)

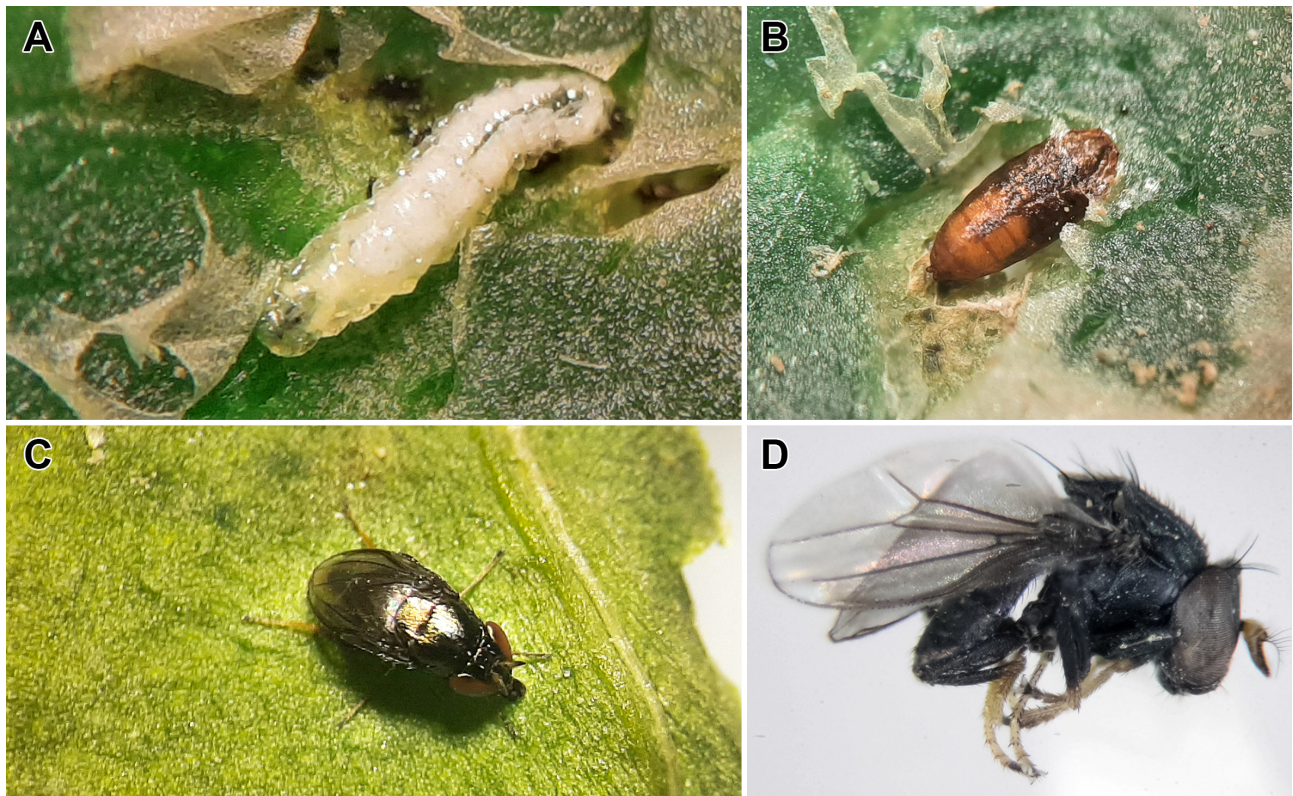
*Discomyza cimiciforme* Haliday, 1855:124.

[Figs 2, 3]

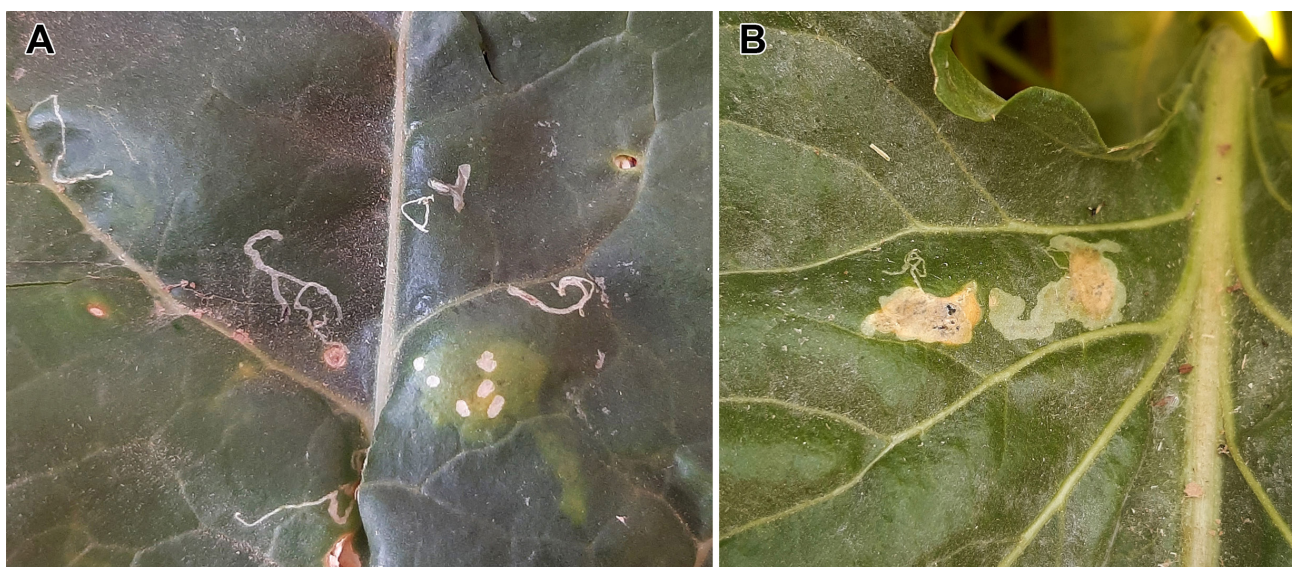
**Diagnosis** (Fig. 2C–D). Body length 1.50–2.35 mm; generally black with shiny metallic reflections ranging from green to copper; head in lateral view two times as long as wide, with a flattened anterior surface; eye highly elongated, ranging in orientation from oblique to almost vertical; interfrons, fronto-orbital plate and face distinctly wrinkled; scape and apex of postpedicel black, other parts of antenna yellowish-orange; arista with 8–11 dorsal setulae; anepisternum strongly wrinkled; wing infuscate, occasionally becoming darker toward the base; halter brownish-black; femora and tibiae, except at their apices, black; tarsi yellow to yellowish-brown; abdominal tergites entirely black, with only sparse microtomentum and exhibit a mostly shiny metallic appearance.

**Material examined.** (Reared from *Beta vulgaris*): 1 ♂, 2 ♀♀ (glued to card point), IRAN: North Khorasan Province, Bojnord, Hesar-e Garmkhan, 37°30'47.4"N 057°29'21.9"E, 5.VIII.2025, 1150 m, reared from *Beta vulgaris*, leg. E. Hamidi [HMIM]. **Other examined material.** 1 ♀, (glued to card point), IRAN: Bushehr Province, Asaluyeh, Nayband, 27°24'45"N 052°40'09"E, 6–8.XI.2021, –10 m, leg. E. Gilasian [HMIM]; 2 ♂♂ 2 ♀♀ (glued to card point), Hormozgan Province, Sirik, Azini Wharf, 26°19'39.9"N 057°06'15.9"E, 13.V.2022, 0 m, leg. M. Mofidi Neyestanak [HMIM]; 3 ♂♂ 2 ♀♀ (glued to card point), Hormozgan Province, Qeshm Island, Kovarzin, 26°48'39"N 055°46'38"E, 3–4.XI.2021, 0 m, leg. E. Gilasian [HMIM].

**Distribution.** **Afrotropical** (Oman, United Arab Emirates, Yemen); **Oriental** (Taiwan); **Palaeartic:** Europe (Austria, Belgium, Bulgaria, Croatia, Cyprus, France, Germany, Great Britain, Greece, Hungary, Ireland, Italy, Malta, Poland, Portugal, Romania, Russia (European Territory), Slovakia, Spain, Turkey); North Africa (Algeria, Canary Islands, Egypt, Morocco, Tunisia); Middle East: Iraq, Israel; Near East: Saudi Arabia (Mathis & Zatwarnicki 1995; Mathis et al. 2017; Dawah et al. 2019).



**Figure 2.** *Clanoneurum cimiciforme* (Haliday). **A.** Larva; **B.** Pupa; **C-D.** Habitus.



**Figure 3.** Damage symptoms on sugar beet leaves caused by *Clanoneurum cimiciforme* (Haliday).



**Figure 4.** Yellow pan traps used in mangrove habitats, Hormozgan Province.

## DISCUSSION

The genus *Clanoneurum* comprises two valid species: *C. cimiciforme* and *C. americanum* Cresson, 1940, which are distributed, respectively, across the Old World, encompassing the western Palaearctic, Afrotropical and Oriental regions (including Taiwan), and the New World, specifically the Nearctic region of North America (Mathis & Zatwarnicki 1995). *Clanoneurum menozzii* Séguy, 1929, and *C. orientale* Hendel, 1913, were proposed as synonyms of *C. cimiciforme* by Canzoneri & Meneghini (1983) and Cresson (1925), respectively, a taxonomic placement that remains valid today.

Based on published reports, the host plants of this genus belong to the family Chenopodiaceae. Genera within this family that have previously been reported as hosts of *C. cimiciforme* include *Atriplex*, *Beta*, *Salicornia*, and *Suaeda* (Enderlein 1936; de Meijere 1947; Pitkin et al. 2019; Kita et al. 2022). Available evidence indicates that leaf-mining damage on sugar beet is attributable not only to *C. cimiciforme* but also to species of *Diasemocera leucostoma* (Meigen, 1830), historically referred to as *Psilopa leucostoma* in the literature. Such incidents have been documented across Europe and North America (Hering 1962; Krämer 1961, 1962; Landis et al. 1967). While both taxa produce superficially similar patterns of foliar injury, the larval tunnels of *C. cimiciforme* exhibit parallel margins and expand in a uniform manner, in contrast to the more irregular and non-uniformly expanding galleries formed by *D. leucostoma* larvae. During the summer of 2025, scattered symptoms of leaf damage consistent with mining activity were observed in sugar beet fields (Fig. 3) in northeastern Iran, specifically in North Khorasan Province. The vermiform larvae of this species are creamy white in color, and the brown pupae are formed within the same leaf mine where larval feeding takes place (Fig. 2A–B). The preliminary drawings of the immature stages of this species were made by de Meijere (1947), who

illustrated the early form of the pupa along with the structure of the anterior spiracles and the larval pharyngeal skeleton with mouth hooks. Following the collection of infested leaves and their maintenance under laboratory conditions, adult specimens were obtained and identified as *C. cimiciforme*. Although field observations currently indicate that the level of damage inflicted by this species on sugar beet is not economically significant, there is a potential risk that its distribution range and impact may increase in the coming years, potentially extending to other sugar beet-growing regions. Therefore, the implementation of monitoring, early detection, and preventive management strategies is considered essential to mitigate the risk of further spread and establishment of this pest within the country.

Specimens of this species had also previously been collected in 2021 and 2022 from mangrove habitats in southern Iran close to the Persian Gulf and Oman Sea using yellow pan traps (Fig. 4). The proximity of the deployed traps to plants belonging to the family Chenopodiaceae in these areas further supports the likelihood of a host association between this species and members of this plant family in those habitats.

#### AUTHOR'S CONTRIBUTION

The authors confirm their contribution to the paper as follows: E. Gilasian: Conceptualization, methodology, investigation, draft preparation, final review and edit, visualization; E. Hamidi: Methodology, investigation, visualization. Both authors read and approved the final version of the manuscript.

#### FUNDING

This research received no specific grant from any funding agencies.

#### AVAILABILITY OF DATA AND MATERIAL

The specimens listed in this study are deposited in the Hayk Mirzayans Insect Museum, Tehran, Iran, and are available from the curator upon request.

#### ETHICS APPROVAL AND CONSENT TO PARTICIPATE

This study only included arthropod material, and all required ethical guidelines for the treatment and use of animals were strictly adhered to in accordance with international, national, and institutional regulations. No human participants were involved in any studies conducted by the authors for this article.

#### CONSENT FOR PUBLICATION

Not applicable.

#### CONFLICT OF INTERESTS

The authors declare that there is no conflict of interest regarding the publication of this paper.

#### GENERATIVE AI STATEMENT

No generative AI tools were used in the preparation of this paper.

#### ACKNOWLEDGMENTS

Our special thanks go to the Iranian Research Institute of Plant Protection for funding this research. The authors sincerely thank Dr. Tadeusz Zatwarnicki (Natural History Museum, Jelenia Góra, Poland) for his assistance in the identification of species and for his constructive comments on the manuscript. We are also indebted to Dr H. Malkeshi and Dr M. Mofidi Neyestanak (Iranian Research Institute of Plant Protection) for their collaboration in collecting the material.

#### REFERENCES

- Canzoneri, S. & Meneghini, D. (1983) Fauna d'Italia XX. *Ephydriidae e Canaceidae*. Calderini, Bologna, I-IX + 337 pp.
- Cresson, E.T. (1925) Studies in the dipterous family Ephydriidae, excluding North and South American faunas. *Transactions of the American Entomological Society*, 51, 227–258.

- Dawah, H.A., Ahmad, S.K., Abdullah, M.A. & Zatwarnicki, T. (2019) An overview of the Ephydriidae (Diptera) of Saudi Arabia. *Zootaxa*, 4711 (3), 401–445. <https://doi.org/10.11646/zootaxa.4711.3.1>
- Enderlein, G. (1936) 22. Ordnung; Zweiflügler, Diptera. Abt. 16, In: Brohmer, P., Ehrmann, P., Ulmer G. (eds) *Die Tierwelt Mitteleuropas*, 6 (2), *Insekten*, Teil III. Quelle & Meyer, Leipzig, 259 pp.
- Hering, E.M. (1962) Neue Blattminenstudien II. Ein neuer Zuckerrüben-Schädling: *Psilopa leucostoma* (Meig) (Dipt., Ephydriidae). *Deutsche Entomologische Zeitschrift*, 9, 53–55. <https://doi.org/10.1002/mmnd.4800090103>
- Hespenheide, H.A. (1991) Bionomics of leaf-mining insects. *Annual Review of Entomology*, 36 (1), 535–560. <https://doi.org/10.1146/annurev.en.36.010191.002535>
- Izzatullayeva, V., Akparov, Z., Babayeva, S., Ojaghi, J. & Abbasov, M. (2014) Efficiency of using RAPD and ISSR markers in evaluation of genetic diversity in sugar beet. *Turkish Journal of Biology*, 38 (4), 429–438. <https://doi.org/10.3906/biy-1312-35>
- Kita, A., Elsayed, A.K. & Tokuda, M. (2022) Intertidal insects associated with halophytic *Suaeda* (Amaranthaceae) in Japan: a case study in Saga, northern Kyushu. *Biodiversity Data Journal*, 10, Article e79184, 11 p. <https://doi.org/10.3897/BDJ.10.e79184>
- Koga, N. (2008) An energy balance under a conventional crop rotation system in northern Japan: Perspectives on fuel ethanol production from sugar beet. *Agriculture, Ecosystems & Environment*, 125 (1–4), 101–110. <https://doi.org/10.1016/j.agee.2007.12.002>
- Krämer, K. (1961) *Psilopa leucostoma* Meigen (Dipt.) als Blattminierer an Zucker- und Futterrüben. *Gesunde Pflanzen*, 13, 264–266.
- Krämer, K. (1962) *Psilopa leucostoma* Mg. (Diptera)- ein Rübenschädling?. *Anzeiger für Schädlingskunde*, 35, 40–42. <https://doi.org/10.1007/BF01865222>
- Landis, B.J., Wallis, R.L. & Redmond, R.D. (1967) *Psilopa leucostoma*, a new leaf miner of sugar beets in the United States. *Journal of Economic Entomology*, 60, 115–118. <https://doi.org/10.1093/jee/60.1.115>
- Meijere, J.C.H. (1947) Over eenige Dipterenlarven, waaronder een galmug, die mijngangen maakt, en twee Dipteren, die gallen op paddenstoelen veroorzaken. *Tijdschrift voor Entomologie*, 88[1945], 49–62.
- Märländer, B., Hoffmann, C., Koch, H.J., Ladewig, E., Merkes, R., Petersen, J. & Stockfisch, N. (2003) Environmental situation and yield performance of the sugar beet crop in Germany: heading for sustainable development. *Journal of Agronomy and Crop Science*, 189 (4), 201–226. <https://doi.org/10.1046/j.1439-037X.2003.00035.x>
- Mathis, W.N. & Zatwarnicki, T. (1995) *A world catalog of the shore flies (Diptera: Ephydriidae)*. Memoirs on Entomology, International. Vol. 4. Associated Publishers, Gainesville, Florida. 423 pp.
- Mathis, W.N. & Zatwarnicki, T. (1998) 3.49. Family Ephydriidae. In: Papp, L. & Darvas, B. (eds) *Contributions to a Manual of Palaearctic Diptera (with special reference to flies of economic importance)*. Vol. 3. Higher Brachycera. Science Herald, Budapest, pp. 537–570.
- Mathis, W.N., Zatwarnicki, T., Stuke, J.H. & Deeming, J.C. (2017) A conspectus on shore-flies from the United Arab Emirates. In: van Harten, A. (Ed.), *Arthropod Fauna of the United Arab Emirates*. Dar Al-Ummah Printing, Publishing, Distribution & Advertising, Abu Dhabi, 6, 636–761.
- Mathis, W.N., Zatwarnicki, T. & Irwin, A.G. (2021) Ephydriidae (Shore flies). In: Kirk-Spriggs, A.H. & Sinclair, B.J. (eds) *Manual of Afrotropical Diptera. Brachycera–Cyclorrhapha, excluding Calyptratae*. Vol. 3. South African National Biodiversity Institute, Pretoria, South Africa, 2193–2231.
- Pitkin, B., Ellis, W., Plant, C. & Edmunds, R. (2019) The leaf and stem mines of British flies and other insects. <http://www.ukflymines.co.uk/index.php> [Accessed February 18, 2026]
- Pour Rahim, R., Najafi, H., Farzadfar, Sh., Ardeh, M.J., Sheikholeslami, M., Fatemy, S., Ghasemi, A. & Arbabi, M. (2016) *Sugarbeet Handbook (Plant Protection)*. Agricultural Research, Education & Extension Organization, Iranian Research Institute of Plant Protection, 157 pp.
- Sharma, S., Kooner, R., Sandhu, S.S., Arora, R., Kaur, T. & Kaur, S. (2017) Seasonal dynamics of insect pests of sugar beet under sub-tropical conditions. *Journal of Agrometeorology*, 19 (1), 81–83. <https://doi.org/10.54386/jam.v19i1.763>
- van Achterberg, C. (2009) Can Townes type Malaise traps be improved? Some recent developments. *Entomologische Berichten*, 69 (4), 129–135.
- Wheeler, A.G.Jr. (1982) *Clanoneurum americanum* (Diptera: Ephydriidae), a leaf miner of the littoral chenopod *Suaeda linearis*. *Proceedings of the Entomological Society of Washington*, 84 (2), 297–300.

## کشف جنس *Clanoneurum* Becker, 1903 (Diptera: Ephydriidae) در ایران، بر اساس گزارش گونه *Clanoneurum cimiciforme* (Haliday)، به عنوان آفت نوظهور چغندر قند

ابراهیم گیلاسیان<sup>۱</sup>، الناز حمیدی<sup>۲</sup>

۱ بخش تحقیقات رده‌بندی حشرات، موسسه تحقیقات گیاهپزشکی ایران، سازمان تحقیقات، آموزش و ترویج کشاورزی، تهران، ایران.  
۲ بخش گیاهپزشکی، مرکز تحقیقات کشاورزی و منابع طبیعی خراسان شمالی، سازمان تحقیقات، آموزش و ترویج کشاورزی، بجنورد، ایران

نویسنده مسؤل: ابراهیم گیلاسیان | [gilasian@iripp.ir](mailto:gilasian@iripp.ir)

**چکیده:** جنس *Clanoneurum* Becker, 1903 در حال حاضر شامل دو گونه معتبر است که در مناطق آفروتروپیکال، نئارکتیک، خاورزمین و پالتارکتیک انتشار دارند. این جنس و گونه‌ای از آن به نام *C. cimiciforme* (Haliday, 1855) برای اولین بار از ایران (استان خراسان شمالی) به عنوان مینوز برگ چغندر قند (*Beta vulgaris* L.) گزارش می‌شود. این گونه همچنین از زیستگاه‌های مانگرو در جنوب ایران (استان‌های بوشهر و هرمزگان) نزدیک به خلیج فارس و دریای عمان جمع‌آوری شده است. ویژگی‌های ریخت‌شناسی جنس و گونه، همراه با تصاویر حشرات کامل، مراحل نابالغ و علایم خسارت روی برگ‌های چغندر قند ارائه شد.

ویراستار علمی

Tadeusz Zatwarnicki

دریافت: ۰۴ اسفند ۱۴۰۴

ویرایش: ۰۳ اردیبهشت ۱۴۰۵

پذیرش: ۰۹ اردیبهشت ۱۴۰۵

انتشار: ۲۰ اردیبهشت ۱۴۰۵

**واژگان کلیدی:** چغندر قند، مینوز برگ، زیستگاه مانگرو، رکورد جدید، *Psilopini*، مگس‌های ساحلی